

THE cDNA-DERIVED AMINO ACID SEQUENCE OF CHICK HEAT SHOCK PROTEIN
M_r 90,000 (HSP 90) REVEALS A "DNA LIKE" STRUCTURE: POTENTIAL SITE
OF INTERACTION WITH STEROID RECEPTORS

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SUMMARY: We report cDNA sequence, the complete derived aa sequence, and a predicted secondary structure of the chick hsp 90, a protein which has been found to form complexes with steroid hormone receptors. The modelling of the most negatively charged "region A" indicates that the α -helices of this portion of hsp 90 mimic DNA configuration. We propose that this region can, in absence of hormone, interact with and cap the positively charged DNA-binding domain of steroid receptors. © 1989 Academic Press, Inc.

The heat shock protein of 90kDa (hsp 90) is a cytosol protein of unknown function abundant at physiological temperature. Changes of its expression during differentiation as well as post-translational modifications have been described (1,2,3,4). Hsp 90 interacts with and may modulate the function of other proteins such as actin (5), heme-regulated eIF2 α kinase (6), tyrosine kinase oncogene products (7) and steroid hormone receptors (8-11).

Steroid receptors are transcriptional regulators which display a common structural organization (12). Two forms have been described for each receptor, with sedimentation coefficients of \sim 4S and \sim 8S. The larger 8S "non active" form, observed in absence of hormone does not bind to DNA, contrary to the smaller "transformed" one (4S). The 8S form includes the hormone binding subunit and hsp 90 ; it can be dissociated in high salt medium, suggesting ionic interaction between the two subunits. The released 4S receptor then can bind DNA (13). It has been suggested that this separation also occurs physiologically in the cell, upon steroid hormone binding (14).

We report here the cloning of chick hsp 90 cDNA and the derived aa sequence. The predicted secondary structure of the protein suggests a model

where one of the possible interaction of hsp 90 with steroid receptors occurs between the most negatively charged "region A" of the heat shock protein, which mimicks the DNA configuration, and the positively charged DNA binding region of the receptors.

MATERIALS AND METHODS

Construction and screening of cDNA libraries, cDNA characterization and sequencing. A pUC 8/9 library (15) and a λ gt11 library (16) from chick oviduct were screened with hsp 90 cDNA inserts of the following recombinant plasmids: p801, containing the human hsp 90 cDNA (17); chick p9.11, previously described (9); and p8.14, characterized in this work. Restriction maps were determined by single and double enzyme digestion. DNA fragments analysed on 1% agarose gels in TBE buffer, were transferred to nitrocellulose and hybridized to radioactively labelled cDNA inserts under stringent conditions (18). Overlapping cDNA inserts were subcloned into M13mp18 and M13mp19 vectors, and sequenced on both strands (19).

Predicted secondary structure. Computer stereodrawing of a modelled protein segment. The secondary structure was predicted using a procedure which combines three different methods (20), and has overall accuracy of 65.5% for the prediction of α -helix, β -strand and coil structures.

The modelling was performed on a PS 390 Evans and Sutherland computer with the MANOSK program (21).

RESULTS

Isolation of chick hsp 90 cDNA. Only a 5' portion of the chick hsp 90 cDNA (p9.11) was cloned from the pUC 8/9 library, since the EcoRI cloning site was an internal cDNA site (9). The hsp 90 cDNA human clone p801 (17) recognized a chick mRNA of the same size as that recognized by p9.11.

With the p801 insert, twenty-four recombinant clones were isolated from pUC 8/9 library. Three clones were analysed in detail; they hybridized with a ~ 3kb mRNA, as previously described for p9.11. The restriction maps of the three clones were identical, and one clone (p8.14) was selected for sequencing.

To make sure that the EcoRI site was an unique internal site of the hsp 90 cDNA, we screened a λ gt11 library from estrogen-stimulated chick oviducts with p9.11 and p8.14 cDNA inserts. From the 200 positive clones, two different clones overlapping the EcoRI site were selected. Sequence analysis on both strands confirmed that the EcoRI site was unique in the chick hsp 90 cDNA.

Nucleotide sequence of the chick hsp 90 cDNA and aa sequence. The nucleotide sequence and the derived aa sequence of hsp 90 are shown in Figure 1. A single open reading frame of 2184 nucleotides starts at the initiator codon ATG and extends to the in-frame termination codon TAA. The predicted molecular size of the chicken hsp 90 is 84,024 daltons; the calculated pI of 5.2, is in agreement with the experimental determinations (11). Two potential sites of phosphorylation by cAMP-dependent kinase at aa 210 and 456, two consensus

[illegible]

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ACA TCA CGT ATC GAG GAG GTG GAT TAAACAGCTTTACAGGAACATCATGAATGTTTCCTTGGCTAATATGAATAAGTTATATTTTGTATATTATGAATGTTACCTGCCAAAA
Thr Ser Arg Met Glu Glu Val Asp
C C H H H R C C
AAAAACAAAATCTTTGACACTTTTGTCTGCATTCCTCTTTATATTTATTTTCAAGATGTTATACCTTTATTTTGTACATTGCTTTTTCAGCTGATGTGAGATACAAATGCCATT
GAGGAGATATTTCTTTAACTGTACAACTTACAGGCAAGTAAGGAGTAGTTATTTTGTCTGTATTAGCTGGTTTGCAGGTGGCAGGGTTTTGACTTATTCTTAATTTGCCAGA
AAGTAACAAAGACGGTATGACATCTGGGTGTTGAGTTTGTAGTCTTATACAGCTCTTCAACTCTGAATATGTCTAACATAGAGTACCTAGTAACTAGGTATCTATGAGAGCTGAC
ACTTGGGAAGCTTTAAACCTTTTGTCTCTAACAAAGGTCTTGATGTTTAAAGTTGTTTTTATACCTGTTCAAGGATCTGGATTGCACCTACCATAGAATAGAATCCACTGTAAATCTCT
ATTGTGACTTTATGCAAACTGTCATGTACAGTCTCAAACTAGAACTGTAAGAATAAAAGTGTTAAGAATGAAAAAATAAAATGAAAAAATAAAATGAAAAAATAAAATGAAAAA
AAAAA

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Figure 1. Nucleotide and deduced amino acid sequences of chick hsp 90. The protein sequence of 728 amino acids resulting from translation of the 2184 base pair open reading frame is shown beneath the DNA sequence. We sequenced across all subcloning restriction sites. Letters of the third line correspond to the predicted secondary structure (20): H = α helix; C = coil; E = β strand.

sequences for casein kinase II at aa 230 and 259, and four potential sites for N-linked glycosylation at positions 50, 287, 393 and 618 are observed. Hsp 90 phosphorylation has been documented, but no glycosylation has been found so far (3).

The size of the cDNA, ~ 2.9kb, accounts almost completely for the length of the messenger of ~ 3kb detected by Northern blot analysis (9).

The deduced aa chick hsp 90 sequence was compared with the homologous proteins of human (22), mouse (23), drosophila (24), *Trypanosoma cruzi* (*T. cruzi*) (25), yeast (26), and *E. coli* (27). The highest aa identity is observed with human (86%), mouse (87%), and drosophila (80%); with yeast and *T. cruzi*, the aa identity is of 64% and 66%, respectively. A significant level of identity (44%) is still observed with prokaryotes (*E. coli*). Two homologous glucose-regulated proteins also are related to the hsp 90 family (28). The human hsp 90 α sequence (29 and E. Hickey personal communication) derived from the p801 plasmid (17), used as a probe in this work, exhibits a higher (96%) identity with the chick hsp 90 sequence than the human hsp 90 β (22). In addition, near to the N-terminal, the QTQDP sequence is present in chick, human α , rabbit (6), and in one of the two mouse hsp 90 (30).

Hsp 90 secondary structure prediction. Modelling of the negatively charged A region. Hsp 90 contains two highly charged regions: A between aa 221 and 290, and B between aa 530 and 581, probably exposed to the solvent and involved in protein-protein interaction. They form mostly α -helices, likely stabilized by Glu/Asp and Lys interactions (31). These two regions delineate three separate globular domains where α -helices largely predominates over the β -sheet structures. The secondary structure prediction is shown in Figure 1 (third line).

The dissociation of hsp 90 from the heterooligomeric 8S form of steroid receptors results in a "acidophilic" activation of the hormone binding molecule (32) which acquires DNA-binding properties. Looking for a polyacidic

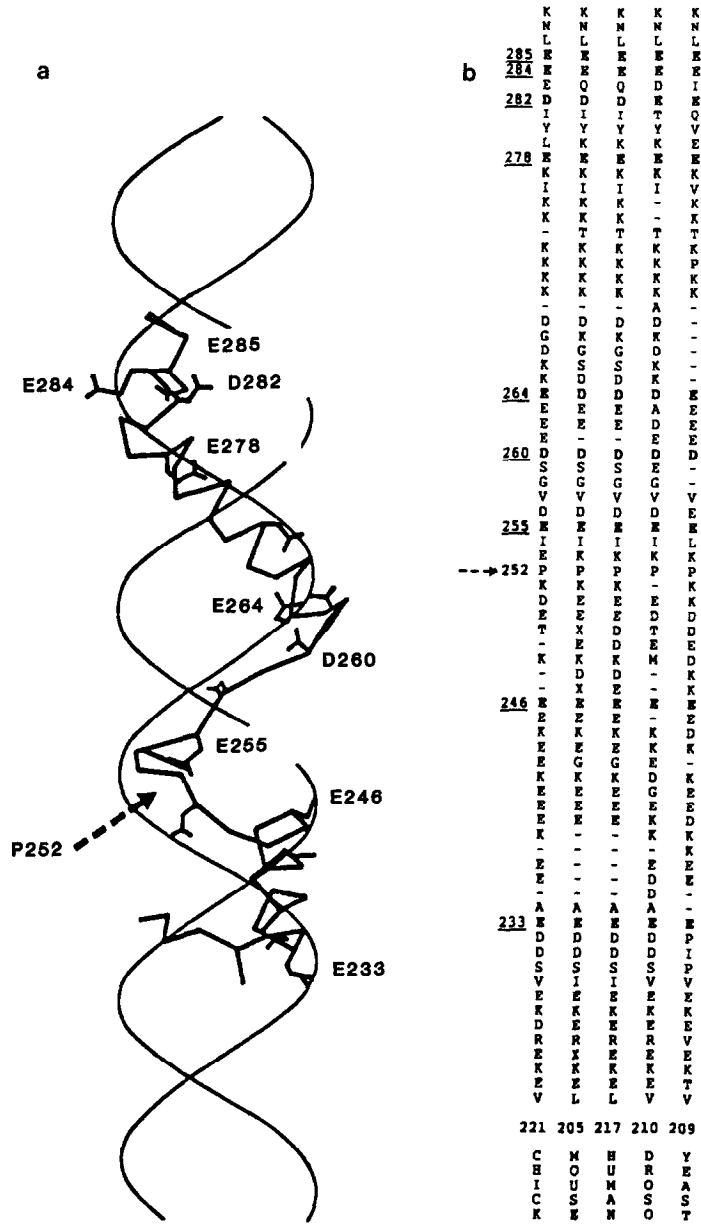


Figure 2. a. Hand drawing of the modelled segment 228-290 of chick hsp 90 A region. The N-terminal segment 221-227, not predicted in α -helix, is not represented. Segment 228-232, predicted with a low probability as helical, was modelled as a coil. The phi and psi angles of the loop region (aa 248-260) were manually adjusted in order to obtain the fitting of a maximum of negatively charged residues to the phosphate backbone of B-DNA structure. During the fitting, the two α -helices (aa 233-247 and aa 261-287) were considered as rigid bodies. Only the α -carbon of the polypeptide backbone and the negatively charged side-chains are represented. b. The numbered residues are the nine most conserved negative amino acids from human to yeast.

stretch of aa which could be involved in hsp 90 receptor interaction, we studied region A which has a large excess of negatively charges aas, mostly Glu. A remarkable topological conservation of the charges in the A region is

observed from human to *Drosophila*, even though the percentage of amino acid identity between species is lower than in other hsp 90 regions (Figure 2a). In the A region, an α -helix is predicted between aa 233 and 247 and, after a proline-containing loop of 13 aa, a second α -helix is predicted between aa 261 and 287. Similar prediction can be made for hsp90 of all species with the exception of *E. coli*, where only the first 17 residues of the A region are present and where steroid hormones have not been detected. A long charged α helix of 32 aa has been observed in troponin C by X-ray crystallography (33), thus confirming the previously predicted structure. Modelling of the hsp 90 A region is represented in Figure 2b. The predicted α -helices have been aligned in space along a double helix of B-DNA, superimposing the carboxyl groups of Glu and Asp with the phosphate groups of the DNA backbone, without imposing any constraint to the polypeptide chain predicted structure. Fifteen aa (11 Glu and 4 Asp), out of 30 negatively charged between aa 228 and 290 are aligned with the DNA phosphate groups and nine are conserved from yeast to human. Thus, the A region of hsp 90 may be viewed in space as a DNA-like, polyanionic structure, capable of interaction with the positively charged DNA-binding region of steroid receptors.

DISCUSSION

We report the isolation and the sequence of chick hsp 90 cDNA ; it contains a single open reading frame encoding a protein with a predicted mass of 84,024 Da, a 5' non-coding segment of 19 nt, and a 3' non-coding segment of 632 nt, in agreement with an almost full length cDNA.

Comparison of aa sequence of chick hsp 90 with that of homologous proteins from *E. coli* to human shows that the hsp 90 family is as highly conserved as the hsp 70 family (1). The chick hsp 90 described here appears to belong to the hsp 90 α -group which is characterized in vertebrates by the sequence QTQDP near to N terminal (6,30).

The charged regions of protein are candidates for ionic interactions with functionally important macromolecules. In hsp 90, the clustering of acidic aa suggests the possibility of salt bridges with basic aa present in the DNA binding region of steroid receptors. Modelling of hsp 90 A region mimicks DNA configuration, and thus support the hypothesis that, in the 8S form of the receptor, hsp 90 masks the DNA binding domain. Experimental data are in agreement with this hypothesis : the 8S form is reversibly dissociated by salts, and the Zn^{2+} is needed for maintaining both DNA and hsp 90 binding property of the receptor (34,35,36). Antibodies raised against hsp 90 A region or receptor DNA binding region recognised their respective antigen, but not the 8S receptor form (37,38). Experiments with glucocorticosteroid receptor indicate that the steroid binding region also is involved in the formation of

the 8S complex (39). This other interaction may facilitate the ionic binding described here, which we believe determinant in inhibiting the receptor function in absence of hormone, by capping the DNA binding region of steroid receptors.

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